

**Table I.** Representative  $^{13}\text{C}$  and  $^1\text{H}$  Spin Systems Identified on the Basis of Two-Dimensional  $^{13}\text{C}$ - $^{13}\text{C}$  and  $^1\text{H}$ - $^{13}\text{C}$  Connectivities

group <sup>a</sup>	carbon atom	chemical shifts <sup>b</sup> (ppm)	
		$^{13}\text{C}$ ( $\pm 0.1$ )	attached $^1\text{H}$ ( $\pm 0.02$ )
ribose	1'	52.2	
	2'	71.2	2.79
	3'	74.6	3.60
	4'	71.0	4.21
	5'	63.8	
isoalloxazine ring	5a	139.4	
	6	129.8	
	7	141.9	
	7a	20.3	
	8	152.8	
	8a	23.0	
alanine-A	o	172.8	
	$\alpha$	51.2	
	$\beta$	21.4	
tyrosine-A	o	171.8	
	$\alpha$	52.3	
	$\beta$	34.9	
	$\gamma$	127.7	
	$\delta$	131.4 <sup>c</sup>	
	$\epsilon$	116.0 <sup>c</sup>	
threonine-A	o	170.8	
	$\alpha$	59.9	
	$\beta$	66.3	
	$\gamma$	16.4	

<sup>a</sup>Sequence-specific assignments have not been made yet for the amino acid spin systems. <sup>b</sup> $^{13}\text{C}$  chemical shifts are relative to TMS. <sup>c</sup> $^1\text{H}$  chemical shifts are relative to TSP. <sup>d</sup>The two tyrosine  $^{13}\text{C}_\delta$  and  $^{13}\text{C}_\epsilon$  carbons appear to have degenerate chemical shifts.

At least 154 of the expected  $\sim 210$   $^{13}\text{C}_\alpha$ - $^{13}\text{C}_\alpha$  correlations were resolved by using the software package MADNMR.<sup>2</sup> This suggests that uniform  $^{13}\text{C}$  labeling will support a heteronuclear approach to sequence-specific resonance assignments. The  $^{13}\text{C}$ - $^{13}\text{C}$  correlations, in combination with multiple-bond  $^{13}\text{C}$ - $^1\text{H}$  correlations or  $^{13}\text{C}$ - $^{15}\text{N}$  correlations from dual  $^{13}\text{C}/^{15}\text{N}$ -labeled proteins, or both, can be used to trace out the peptide backbone connectivities.<sup>1,10,11</sup>

Sensitivity considerations limit the application of the  $^{13}\text{C}$ - $^{13}\text{C}$  DQC experiment to proteins enriched with  $^{13}\text{C}$ . Current methods for incorporating stable isotopes into biotechnology derived proteins have begun to alleviate this problem.<sup>12</sup> Carbon-13 enrichment levels of 20-30% represent a good compromise between improved sensitivity and decreased spectral simplicity. Higher enrichment levels might be useful for providing long-range carbon-carbon coupling constants for selectively enriched proteins<sup>1</sup> but would result in increased spectral overlap in a uniformly enriched protein.

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## First Direct Observation of Pyridyne: Matrix Infrared Study of the Photolysis Products of 3,4-Pyridine Dicarboxylic Anhydride

H.-H. Nam and G. E. Leroi\*

Department of Chemistry, Michigan State University  
East Lansing, Michigan 48824

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Heteroarynes have been proposed as likely intermediates in many organic reactions, principally those involving cycloaddition or cine-substitution.<sup>1</sup> However, only indirect evidence, based on trapping experiments to verify the presence of heteroaryne intermediacy, has been obtained. The reliability of such inferences is severely limited. Other mechanisms, e.g., addition-elimination, trans-halogenation, or addition ring opening-elimination ring closure (ANRORC), also can account for the formation of observed products.<sup>1</sup> Mass spectrometric analysis following the electron impact or the pyrolytic fragmentation of several heteroaryne dicarboxylic anhydrides has been used to conjecture the structure of heteroarynes corresponding to certain  $m/z$  peaks.<sup>2-5</sup> Although diazabiphenylene, the dimer of 3,4-pyridyne, has been identified in the time of flight mass spectrometric and kinetic UV spectroscopic analysis of the products formed by flash photolysis of pyridine-3-diazonium-4-carboxylate,<sup>6</sup> no direct observation of any heteroaryne has yet been published.

In this report we present the first infrared spectrum of 3,4-pyridyne (3,4-didehydropyridine), generated via near UV photolysis ( $\lambda > 340$  nm) of 3,4-pyridine dicarboxylic anhydride (3,4-PDA) in  $\text{N}_2$  or Ar matrices. Similar experiments by Dunkin and McDonald were not successful;<sup>7</sup> apparently the photolytic conditions utilized in that study produced only decomposition products of the desired heteroaryne.

3,4-PDA (obtained from Aldrich and vacuum sublimed before use) was sublimed and codeposited for 2 h with Ar or  $\text{N}_2$  (flow rate 2 mmol/min) on the CsI substrate of an Air Products CS202 Displex cryostat. Photolyses were conducted with a 200 W Hg-Xe arc lamp equipped with a water filter and various cutoff filters. Infrared spectra of the precursor and photolyzed products at 13 K were recorded with a BOMEM DA3.01 interferometric spectrometer.

As summarized in Scheme I, mild irradiation ( $\lambda > 340$  nm and less than 100 min duration) of 3,4-PDA in  $\text{N}_2$  or Ar matrices at 13 K readily fragmented the precursor to form CO,  $\text{CO}_2$ , and 3,4-pyridyne, which has a strong peak at  $2085\text{ cm}^{-1}$  diagnostic of carbon-carbon triple bond formation. Subsequent irradiation with  $\lambda > 210$ -nm light immediately decomposed 3,4-pyridyne into HCN, diacetylene, acetylene, and cyanoacetylene as a result of alternative two-bond scissions. The infrared spectrum in the  $2050$ - $2300\text{ cm}^{-1}$  region prior to and following controlled photolysis (Figure 1) clearly demonstrates the formation of 3,4-pyridyne and its subsequent decomposition. The peak due to 3,4-pyridyne at  $2085\text{ cm}^{-1}$  disappears upon shorter wavelength irradiation, and new peaks at  $2101\text{ cm}^{-1}$  (HCN),  $2181\text{ cm}^{-1}$  (diacetylene) and  $2236\text{ cm}^{-1}$  (cyanoacetylene) begin to grow. Ten additional peaks below  $2000\text{ cm}^{-1}$  show the same growth and decay pattern as the  $2085\text{-cm}^{-1}$  band and are also attributable to 3,4-pyridyne (Table I).

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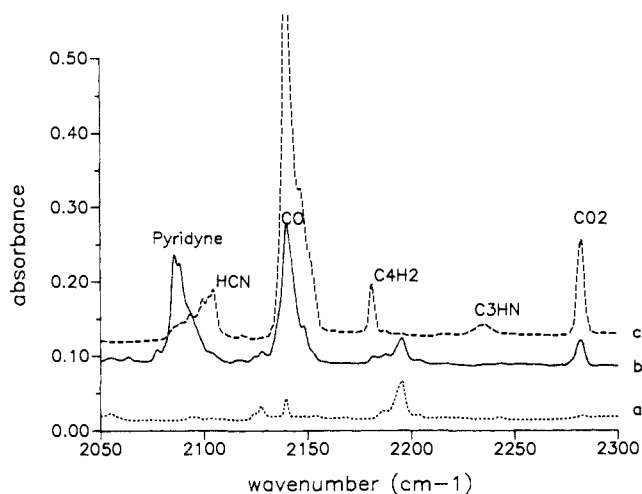
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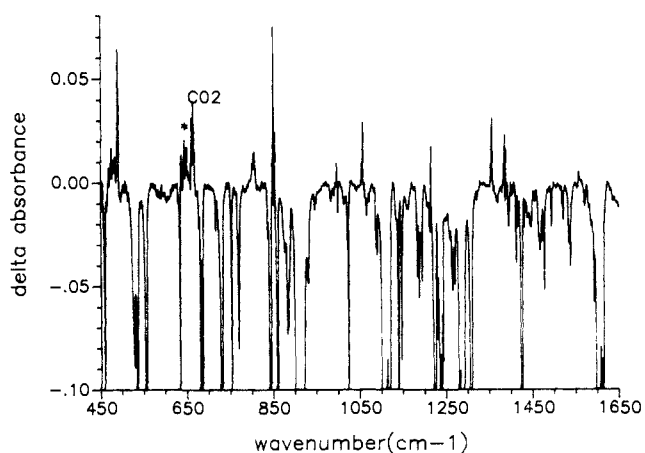
**Figure 1.** IR spectra of 3,4-PDA and its photolyzed products in the 2050–2300-cm<sup>-1</sup> region in an N<sub>2</sub> matrix at 13 K: (a) 3,4-PDA; (b) after 100 min photolysis through water and λ > 340-nm filter (The peak at 2281 cm<sup>-1</sup> is due to <sup>13</sup>CO<sub>2</sub>); (c) following additional 30 min photolysis with λ > 210 nm.

**Table I.** Infrared Bands (cm<sup>-1</sup>) Resulting from Photolysis of 3,4-PDA in an N<sub>2</sub> Matrix at 13 K<sup>d</sup>

λ > 340 nm <sup>a</sup>	λ > 210 nm <sup>b</sup>	photolyzed products	<i>o</i> -benzyne <sup>c</sup>
	2236	cyanoacetylene	
	2181	diacetylene	
	2101	HCN	
2085		3,4-pyridyne	2082
1558		3,4-pyridyne	1596
			1448
1387		3,4-pyridyne	1395
1355		3,4-pyridyne	1355
	1260	polymer	
1216		3,4-pyridyne	
1055		3,4-pyridyne	1055
			1038
996		3,4-pyridyne	
853		3,4-pyridyne	
848		3,4-pyridyne	848
802		3,4-pyridyne	
	751	acetylene	
744	744	acetylene	
			739
	703	polymer	
	673	cyanoacetylene	
648	648	diacetylene	
635	635	diacetylene	
489		3,4-pyridyne	470

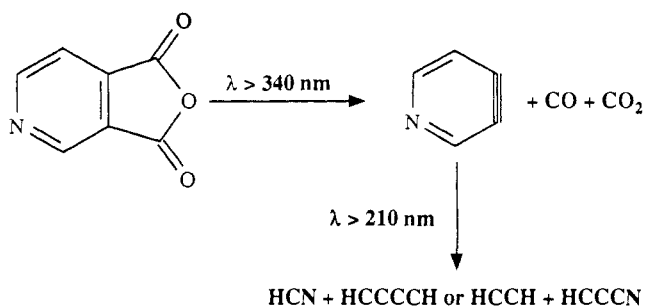
<sup>a</sup>Photolysis of 3,4-PDA. (100 min). <sup>b</sup>Additional 30 min photolysis after a. <sup>c</sup>Reference 9. <sup>d</sup>Comparison to *o*-benzyne (last column).

The IR frequencies of 3,4-pyridyne indicate that this molecule is remarkably similar to *o*-benzyne in character. The wavenumbers observed for both *o*-benzyne and 3,4-pyridyne in N<sub>2</sub> matrices are collected in Table I. However, 3,4-pyridyne decomposes much



**Figure 2.** Difference spectrum of 3,4-PDA before and after mild photolysis. \* indicates a band due to diacetylene.

**Scheme I**



faster. Although crude thermodynamic calculations suggest similar ring strain energy for these two molecules (~60 kcal/mol),<sup>8</sup> 3,4-pyridyne has less resonance energy, which may account for its lower stability.

Unlike 3,4-pyridyne, the 2,3-isomer could not be isolated under our experimental conditions. Additional experiments to identify the products of 2,3-PDA photolysis are in progress. The results, plus theoretical calculations of the structures and vibrational frequencies of various heteroarynes, will be reported in a future publication.

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